

# Kinetics and mechanism of large rate enhancement in an acidic aqueous cleavage of the tertiary amide bond of *N*-(2-methoxyphenyl)-*N'*-morpholinophthalamide (**1**)

Yoke-Leng Sim, Azhar Ariffin, M. Niyaz Khan\*

Department of Chemistry, Faculty of Science, University of Malaya, 50603 Kuala Lumpur, Malaysia

## ARTICLE INFO

### Article history:

Received 31 January 2008

Available online 25 April 2008

### Keywords:

*N*-(2-Methoxyphenyl)-*N'*-

morpholinophthalamide

*N*-(2-Methoxyphenyl)phthalimide

Acidic hydrolysis

Kinetics

Intramolecular rate enhancement

Reaction mechanism

## ABSTRACT

The rate of conversion of **1** to *N*-(2-methoxyphenyl)phthalimide (**2**) within [HCl] range  $5.0 \times 10^{-3}$ –1.0 M at 1.0 M ionic strength (by NaCl) reveals the presence of both uncatalyzed and specific acid-catalyzed kinetic terms in the rate law. Intramolecular carboxamide group-assisted cleavage of amide bond of **1** reveals rate enhancement of much larger than  $10^6$ -fold compared to the expected rate of analogous intermolecular reaction.

© 2008 Elsevier Inc. All rights reserved.

## 1. Introduction

The classical paper of Shafer and Morawetz revealed more than  $10^6$ -fold rate enhancement in the specific base-catalyzed carboxamide group-assisted cleavage of amide bond of phthalamide and *N,N'*-dimethylphthalamide in aqueous solvent [1]. Since the huge rate enhancements due to intramolecularity of reactions bear a striking resemblance to enzyme-catalyzed reactions [2] a surge of interest in this area of research can be seen in the literature [2d,2e,3]. Almost all such and closely related studies involve cyclization coupled with only hydroxide ion and general base catalysis [4]. A search of literature reveals only one report on intramolecular aminolysis of amides where the cyclization of 2-aminomethylbenzamide to phthalimidine has been also studied within the pH 5 to 5 M HCl [5]. But the kinetic data analysis for data obtained within the pH 5 to 5 M HCl was complicated due to the presence of rather more basic 2-aminomethyl group in the substrate containing the amide group. Cohen and Lipowitz [6] reported acid-catalyzed hydrolysis of *o*-benzamido-*N,N*-dicyclohexylbenzamide where neighboring amide group assistance turned out to be  $>10^4$ -fold. These authors followed a synthetic approach and consequently a quantitative estimation of neighboring amide group assistance as well as fine details of mechanism remained unclear. Nearly  $10^4$ - to  $10^{14}$ -fold rate enhancement has been reported in the intramolecular carboxyl group-assisted cleavage of amide bond at pH  $\leq 3$  [7]. The aim of the present study was to explore quantita-

tively the probable rate enhancement in the intramolecular amide group-assisted cleavage of another neighboring amide bond of **1** at pH  $< 3$ . The observed results and their probable explanation(s) are described in this manuscript.

## 2. Experimental

### 2.1. Materials

Synthesis of *N*-(2-methoxyphenyl)phthalimide (**2**) is described elsewhere [8]. Synthesis of *N*-(2-methoxyphenyl)-*N'*-morpholinophthalamide (**1**), *N*-benzoylmorpholine (**3**), kinetic measurements and product identification are described in [Supplementary Data \(SD\)](#). Stock solutions of **1** (0.025 M) and **3** (0.01 M) were prepared in acetonitrile.

## 3. Results and discussion

### 3.1. Aqueous cleavage of **1** at different [HCl] and 35 °C

The rate of aqueous cleavage of **1** was studied within [HCl] range  $5.0 \times 10^{-3}$ –1.0 M at constant ionic strength of 1.0 M (by NaCl). It is noteworthy that light precipitates appeared into the reaction mixtures after reaction time  $t \geq 4.7$  h (i.e.  $\geq \sim 1$  halflife) at  $\leq 0.05$  M HCl. But this kinetic problem is solved as described in the 'Kinetic Measurements' in [Supplementary Data \(SD\)](#). Similar observations were obtained into the reaction mixture for acid-catalyzed hydrolysis of **2** under similar conditions [8].

\* Corresponding author. Fax: +60 3 79674193.

E-mail address: [niyaz@um.edu.my](mailto:niyaz@um.edu.my) (M.N. Khan).

Pseudo-first-order rate constants ( $k_{\text{obs}}$ ) at different [HCl] fit to the following empirical equation

$$k_{\text{obs}} = \frac{k_0 + k_c K [\text{HCl}]}{1 + K [\text{HCl}]} \quad (1)$$

where  $k_0$ ,  $k_c$  and  $K$  represent empirical constants. The nonlinear least-squares calculated values of  $k_0$ ,  $k_c$  and  $K$  are summarized in Table 1. The extent of the reliability of observed data fit to Eq. (1) is evident from the plot of Fig. 1 where solid line is drawn through the least-squares calculated data points. The absolute percent residual errors  $\{\text{ARE} = 100 \times |(k_{\text{obsi}} - k_{\text{calcdi}})/k_{\text{obsi}}|\}$  where  $k_{\text{obsi}}$  and  $k_{\text{calcdi}}$  represent respective observed and calculated values of pseudo-first-order rate constants at the  $i$ th value of [HCl] are  $\leq 2\%$  at  $\geq 0.01$  M HCl while ARE = 5–6% at  $5.0 \times 10^{-3}$  M HCl.

The standard deviation (=36%) associated with  $k_0$  is significantly large and consequently a skeptic might think that  $k_0$  is not significantly different from zero. Although the standard deviation is large (36%), perhaps it is not too large to believe that  $k_0$  is different from zero for the following calculated results. (i) The contributions of  $k_0$  compared to  $k_c K [\text{H}^+]$  in the numerator of Eq. (1) are 62, 45, 14, 8 and 4% at 0.005, 0.01, 0.05, 0.1 and 0.2 M HCl, respectively. (ii) The attempt to calculate  $k_c$  and  $K$  from Eq. (1) with  $k_0 = 0$  resulted in systematic positive ARE as 59, 42, 13, 3 and < 1% at the respective 0.005, 0.01, 0.05, 0.1 and >0.1 M HCl. But the values of ARE at the corresponding values of [HCl], turned out to be –6, –2, 2, –1 and < 1% when the data treatment with Eq. (1) involved  $k_0$ ,  $k_c$  and  $K$  as three unknown parameters. The results described above as (i) and (ii) demonstrate that  $k_0$  cannot be neglected compared to  $k_c K [\text{H}^+]$  within the [HCl] range of present study.

The downward curvature of the plot of Fig. 1 does not seem to be visible to the naked eye. So a few kinetic runs were carried out within 2.0–5.0 M HCl in the absence of NaCl (i.e. ionic strength was not kept constant) and the  $k_{\text{obs}}$  values, under such conditions, showed upward curvature (Fig. 2) which could be ascribed to the ionic strength effect. Thus, a few kinetic runs were also carried out at constant ionic strength (5.0 M by NaCl) and within [HCl] range 2.0–5.0 M. An attempt to carry out the kinetic run at 1.0 M HCl and 5.0 M ionic strength (by NaCl) was unsuccessful because the reaction mixture became turbid at  $t > \sim 300$  s. The values of  $k_{\text{obs}}$  within [HCl] range 2.0–5.0 M HCl at 5.0 M ionic strength were treated with Eq. (1) considering  $k_0$  as known parameter and the nonlinear least-squares calculated values of  $k_c$  and  $K$  are shown in Table 1, where  $k_0 = 2.19 \times 10^{-5} \text{ s}^{-1}$ . The data fit appears satisfactory as evident from Fig. 2.

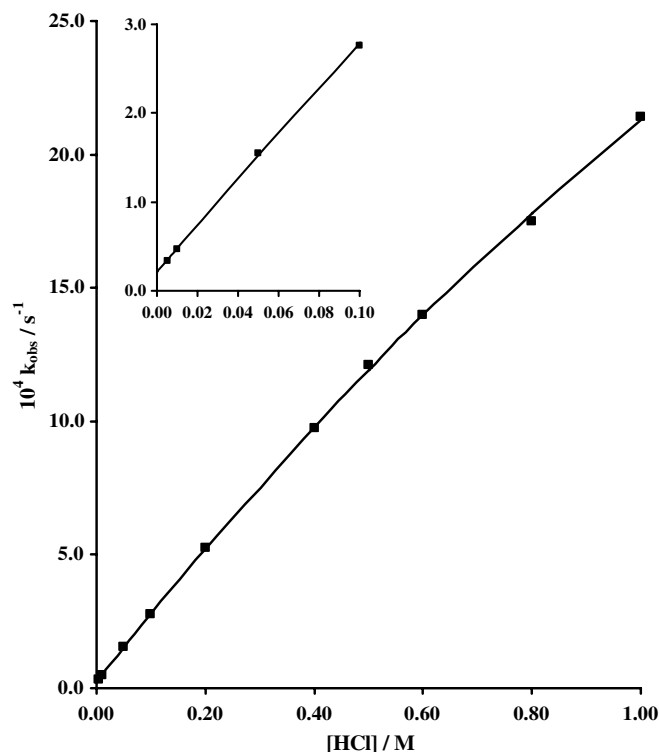


Fig. 1. The plot showing the dependence of pseudo-first-order rate constants,  $k_{\text{obs}}$ , upon [HCl] for acidic hydrolysis of **1** at  $2.0 \times 10^{-4}$  M **1**, 1.0 M ionic strength (by NaCl) and 35 °C. The solid line is drawn through the least-squares calculated data points using Eq. (1).

Unlike NaCl, the use of NaBr to maintain the constant ionic strength at 5.0 M did not produce turbidity problem into the reaction mixture at  $\leq 1.0$  M HCl. Thus, a few kinetic runs were also carried out within [HCl] range 0.2–5.0 M. The values of  $k_{\text{obs}}$  obtained under such conditions, as shown graphically in Fig. 2, were found to fit to Eq. (1) and the nonlinear least-squares calculated values of  $k_0$ ,  $k_c$  and  $K$  are shown in Table 1. Negative value of  $k_0$  with standard deviations of more than 600% merely indicates that the contribution of  $k_0$  compared to  $k_c K [\text{HCl}]$  in the numerator of Eq. (1) is insignificant and hence a relatively more reliable values of  $k_c$  and  $K$  were obtained from Eq. (1) by considering  $k_0 = 21.9 \times 10^{-6} \text{ s}^{-1}$  obtained at 1.0 M ionic strength. Such calculated values of  $k_c$  and  $K$  are also summarized in Table 1.

Table 1  
Values of  $k_0$ ,  $k_c$  and  $K$  calculated from Eq. (2) for the aqueous cleavage of **1** and **3**<sup>a</sup>

Amide	$\mu^b$ (M)	Temp (°C)	$10^7 k_0$ ( $\text{s}^{-1}$ )	$10^4 k_c$ ( $\text{s}^{-1}$ )	$10^2 K$ ( $\text{M}^{-1}$ )	$\text{pK}_a^c$	[HCl] range (M)	# of runs
<b>1</b>	1.0	35	$219 \pm 78^d$	$104 \pm 9^d$	$25.3 \pm 2.8^d$	$-0.60$ ( $-0.75$ ) <sup>e</sup>	0.005–1.0	10
	5.0	35	219	$262 \pm 17$	$33.3 \pm 4.7$	–0.48	2.0–5.0	4
	5.0 <sup>f</sup>	35	$-740 \pm 4640$	$227 \pm 11$	$46.9 \pm 6.8$	–0.33	0.2–5.0	8
			219	$229 \pm 7$	$45.7 \pm 3.3$	–0.34	0.2–5.0	8
<b>3</b>	1.0 <sup>b</sup>	65	$0.36 \pm 3.54$	$0.165 \pm 0.060$	$66.7 \pm 39.7$	–0.18	0.05–1.0	6
		65	0	$0.161 \pm 0.036$	$69.8 \pm 23.5$	–0.16	0.05–1.0	6
		35 <sup>g</sup>	0	$2.81 \times 10^{-3}$				
Benzamide <sup>h</sup>	6.0 <sup>i</sup>	25	0	$(8.34 \pm 0.28) \times 10^{-3}$	$81.0 \pm 9.7$	–0.09	1.0–6.0	6

<sup>a</sup>  $[\text{1}]_0 = 2.0 \times 10^{-4}$  M,  $\lambda = 290$  nm, aqueous reaction mixture for each kinetic run contained 0.8% v/v  $\text{CH}_3\text{CN}$ .  $[\text{3}]_0 = 1.3 \times 10^{-4}$  M,  $\lambda = 230$  nm, aqueous reaction mixture for each kinetic run contained 1.3% v/v  $\text{CH}_3\text{CN}$ .

<sup>b</sup> Ionic strength was maintained by NaCl.

<sup>c</sup>  $\text{pK}_a = \log K$ .

<sup>d</sup> Error limits are standard deviations.

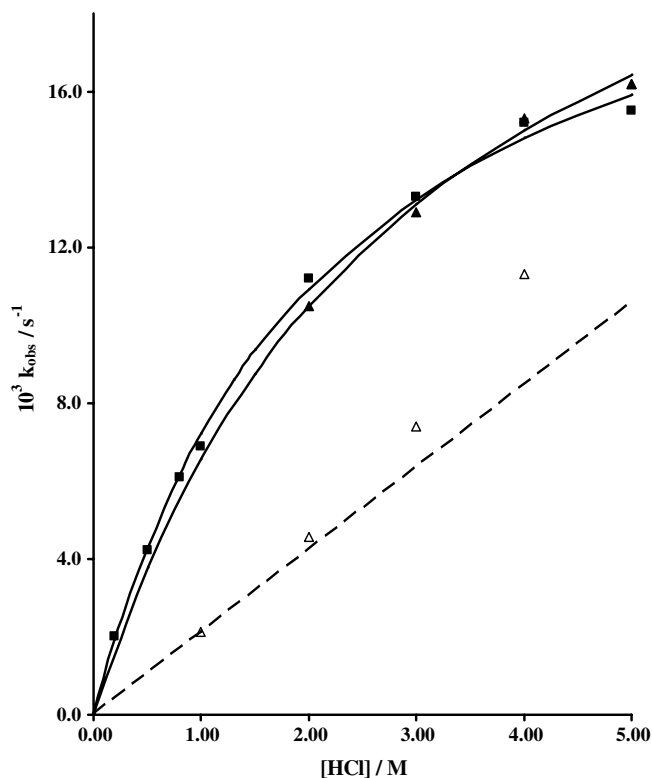
<sup>e</sup> Thermodynamic  $\text{pK}_a$  (=concentration  $\text{pK}_a + \log \gamma$  where activity coefficient  $\gamma = 0.70$  at 1.0 M ionic strength).

<sup>f</sup> Ionic strength was kept constant by NaBr.

<sup>g</sup> The value of  $k_c$  at 35 °C was estimated from  $k_c$  value at 65 °C using Eyring equation.

<sup>h</sup> The observed data ( $k_{\text{obs}}$  versus [HCl]) were obtained from Ref. [9].

<sup>i</sup> Ionic strength was kept constant by LiCl.



**Fig. 2.** The plots showing the dependence of pseudo-first-order rate constants,  $k_{\text{obs}}$ , upon  $[\text{HCl}]$  for aqueous cleavage of **1** at  $2.0 \times 10^{-4}$  M **1**, 5.0 M ionic strength (by NaBr (■) and by NaCl (▲)) and 35 °C. The solid lines are drawn through the least-squares calculated data points using Eq. (1). The symbol (Δ) represents  $k_{\text{obs}}$  obtained in the absence of NaCl, i.e. ionic strength was not kept constant in these kinetic runs.

It may be noted that the values of  $k_{\text{obs}}$ , obtained within  $[\text{HCl}]$  range 2.0–5.0 M 5.0 M ionic strength, vary by < 7% in the presence of NaCl and NaBr. However, the observed data fit to Eq. (1) at 5.0 M ionic strength by NaCl is considered to be less reliable compared to that at 5.0 M ionic strength by NaBr because of the lack of  $k_{\text{obs}}$  value at  $\leq 1.0$  M HCl in the presence of NaCl.

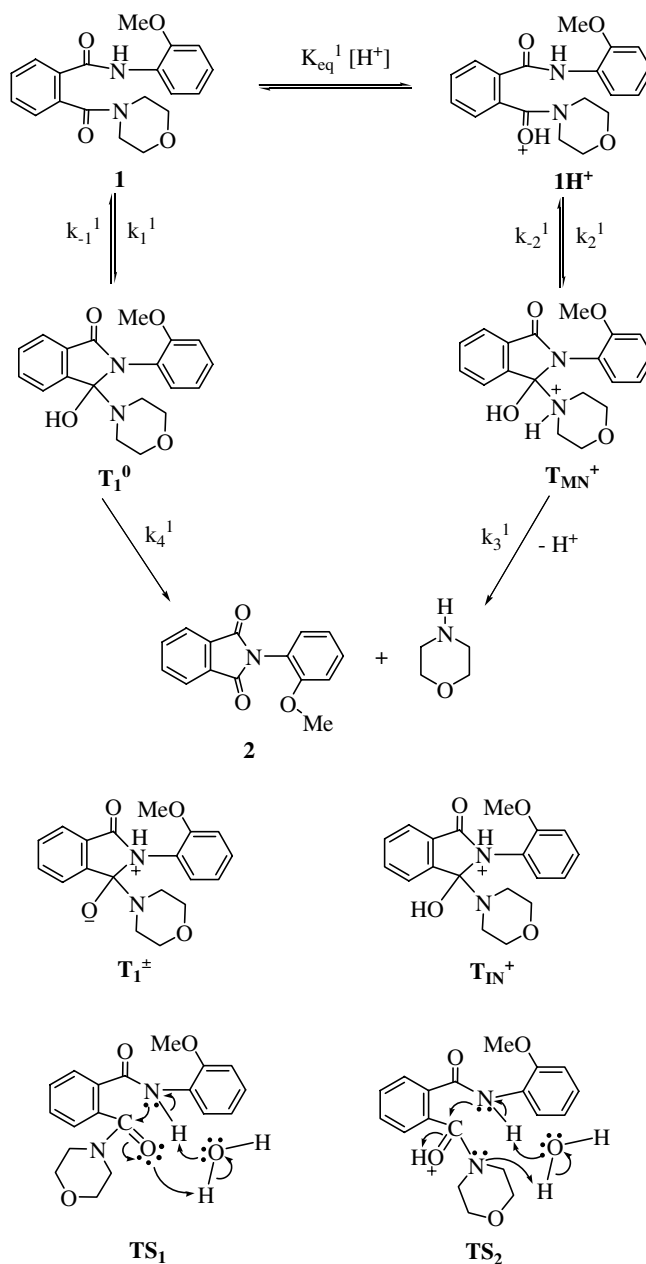
### 3.2. Aqueous cleavage of *N*-benzoylmorpholine (**3**) at different $[\text{HCl}]$ and 65 °C

Rate of specific acid-catalyzed hydrolysis of **3** was studied within  $[\text{HCl}]$  range 0.05–1.0 M at 1.0 M ionic strength (by NaCl). Pseudo-first-order rate constants ( $k_{\text{obs}}$ ), as shown graphically in Fig. 1 (Supplementary Data, SD), showed reasonably good fit to Eq. (1). The nonlinear least-squares calculated values of  $k_0$ ,  $k_c$  and  $K$  are summarized in Table 1. The calculated value of  $k_0$  is associated with extremely high (~10-fold) standard deviation (Table 1) which shows insignificant contribution of  $k_0$  compared to  $k_c K[\text{HCl}]$  in Eq. (1) and consequently the calculated value of  $k_0$  is unreliable. Thus, the values of  $k_c$  and  $K$  were also calculated from Eq. (1) with  $k_0 = 0$  and such calculated values of  $k_c$  and  $K$  are shown in Table 1. It is evident from Table 1 that the values of  $k_c$  and  $K$  remained almost unchanged with change in  $k_0$  value from 0.0 to  $3.6 \times 10^{-8} \text{ s}^{-1}$ . The  $k_0$ -step could not be detected in the specific acid-catalyzed hydrolysis of various amides [9,10]. The value of  $k_{\text{obs}}$  ( $=1.39 \times 10^{-8} \text{ s}^{-1}$ ) at 0.1 M HCl and 1.0 M ionic strength may be compared with  $k_{\text{obs}} = 1.7 \times 10^{-9} \text{ s}^{-1}$  obtained for hydrolysis of *N,N*-dimethylbenzamide at 0.12 M HCl and 0.12 M ionic strength and 35 °C [9]. The value of pseudo-first-order rate constant ( $k_0$ ) for neutral uncatalyzed hydrolysis of formamide is  $6 \times 10^{-10} \text{ s}^{-1}$  (at 35 °C) [11].

The values of  $k_{\text{obs}}$ , obtained for hydrolysis of benzamide within  $[\text{HCl}]$  range 1.0–6.0 M at 25 °C and constant ionic strength 6.0 M (by LiCl) [9], were found to fit reasonably well to Eq. (1) with  $k_0 = 0$  and the nonlinear least-squares calculated values of  $k_c$  and  $K$  are shown in Table 1. The effects of  $[\text{HCl}]$  on  $k_{\text{obs}}$  for hydrolysis of benzamide and *N,N*-dimethylbenzamide reveal ~3-fold larger value of  $k_{\text{obs}}$  for benzamide than that for *N,N*-dimethylbenzamide under similar experimental conditions [9].

Unlike the mechanisms of thoroughly studied highly efficient hydroxide ion-catalyzed intramolecular carboxamide group-assisted cleavage of amide bonds, the mechanisms of rarely studied specific acid (hydronium ion)-catalyzed such reactions are not well explored. However, a plausible mechanism of such a reaction as described in the present manuscript may be shown by Scheme 1 where the protonation of **1** in an acidic medium is shown by an equilibrium process with equilibrium constant  $K_{\text{eq}}^1$ .

In Scheme 1, if the expected immediate intramolecular addition product,  $T_1^\pm$  and  $T_{\text{IN}}^+$  in respective  $k_1^1$ -step and  $k_2^1$ -step are too unsta-



**Scheme 1.** Structures  $T_1^\pm$ ,  $T_{\text{IN}}^+$ ,  $\text{TS}_1$ ,  $\text{TS}_2$ .

ble to exist for a period of longer than  $10^{-13}$  s [12], then the formation of  $T_1^0$  and  $T_{MN}^+$  are expected to occur through transition states  $TS_1$  and  $TS_2$ , respectively. The fact that the  $pK_a$  of conjugate acid of leaving group is larger in  $k_4^1$ -step than that in  $k_{-1}^1$ -step and also the release of five-membered ring strain in  $k_{-1}^1$ -step make  $k_{-1}^1 \gg k_4^1$  and this inequality leads to  $k_4^1$ -step as the rate-determining step. The  $pK_a$  of conjugate acid of leaving group in  $k_3^1$ -step is significantly smaller than that in  $k_{-2}^1$ -step which in turn imply that  $k_3^1 \gg k_{-2}^1$ . But the release of five-membered ring strain in  $k_{-2}^1$ -step makes  $k_{-2}^1 \gg k_3^1$ . Thus, the combined effects of relative  $pK_a$  of conjugate acids of leaving groups in  $k_3^1$ -step and  $k_{-2}^1$ -step as well as release of five-membered ring strain in only  $k_{-2}^1$ -step would dictate whether  $k_3^1$ -step or  $k_{-2}^1$ -step is the rate-determining step. The observed rate law:  $\text{rate} = k_{\text{obs}} [\mathbf{1}]_T$  and Scheme 1 can lead to Eq. (2)

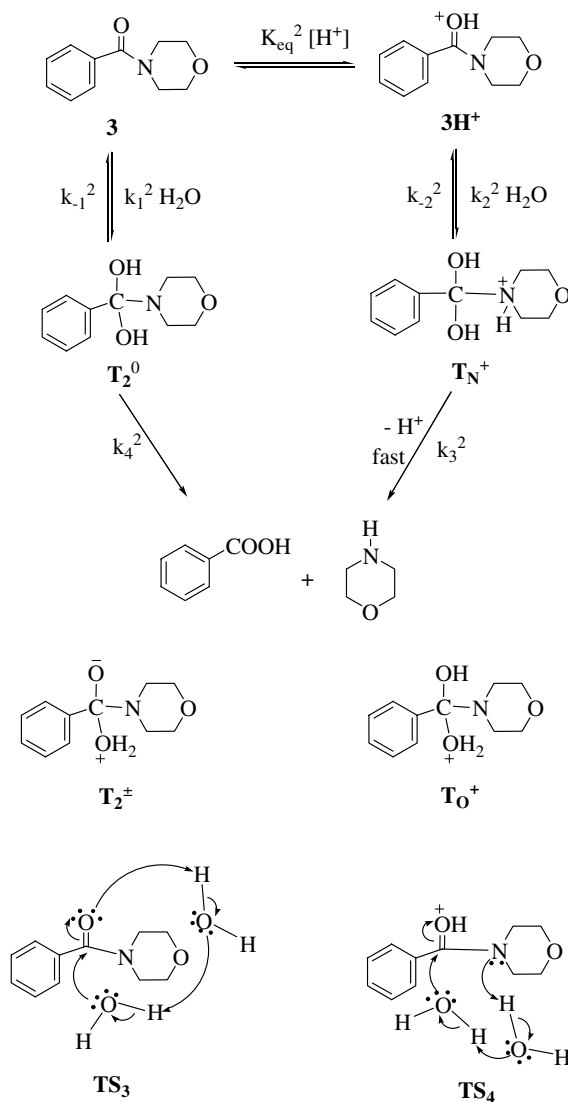
$$k_{\text{obs}} = \frac{k_4^1 K_1^1 + k_3^1 K_2^1 K_{\text{eq}}^1 [\text{H}^+]}{1 + K_{\text{eq}}^1 [\text{H}^+]} \quad (2)$$

where  $K_1^1 = k_1^1/k_{-1}^1$ , and  $K_2^1 = k_2^1/k_{-2}^1$ . Eq. (2) is similar to Eq. (1) with  $k_0 = k_4^1 K_1^1$ ,  $k_c = k_3^1 K_2^1$  and  $K = K_{\text{eq}}^1$ . Thus, the ratio  $k_c/k_0$  ( $\approx 500$ ) shows that specific acid-catalytic factor is  $\sim 500$ -fold in the cyclization of **1** to **2**.

One of the reviewers has raised an interesting point that whether the amide group acts as a nitrogen nucleophile through the enol form—which is the only species with a lone pair of electrons on N available to act as a nucleophile, or not.<sup>1</sup> It is well known that C–N bond in an amide is neither a pure single bond nor a pure double bond. The nucleophilicity of the amide nitrogen is essentially governed by its basicity. The reported  $pK_a$  values of  $\text{MeC}\equiv\text{NH}^+$ ,  $\text{Me}_2\text{C}=\text{NHMe}^+$  and  $\text{MeNH}_3^+$  are  $-10.1$ ,  $5.5$  and  $10.7$ , respectively [13]. The values of  $\sigma_p^{\text{COMe}} = 0.50$ ,  $\sigma_m^{\text{COMe}} = 0.38$ ,  $\sigma_p^{\text{COPh}} = 0.44$ ,  $\sigma_m^{\text{COPh}} = 0.36$ ,  $\sigma_{\text{COMe}}^- = 0.82$  and  $\sigma_R^-(\text{COMe}) = 0.32$  [14]. In view of these findings, it seems difficult to believe that the amide nitrogen is a stronger base in pure enol form (where lone pair of electrons of N exist in  $sp^2$  orbital) than in pure keto form (where  $sp^3$  orbital contains lone pair of electrons of the amide N). Although the kinetic data of the present study are not sufficient to rule out completely the possibility of the  $sp^2$  nitrogen of the enol form of the amide acting as the nucleophile in the conversion of **1** to **2**, the mechanism shown in Scheme 1 is considered to be more appropriate for the following reasons: (i) experimental and theoretical evidence for enol formation in the aqueous solutions of primary and secondary amides are lacking (at least to the best of our literature search) and (ii) the enol form of the amide group has not been considered in any closely related studies [7g,15,16].

The specific acid-catalyzed hydrolysis of amides has been extensively studied [17]. In view of these extensive experimental studies and a recent theoretical study [18], a plausible reaction mechanism for hydrolysis of **3** in an acidic aqueous solution is shown in Scheme 2 where the catalytic reaction involves  $k_2^2$ -step as the rate-determining step [17–19]. In Scheme 2, if the immediate intermolecular addition intermediates  $T_2^\pm$  and  $T_0^+$  in respective  $k_1^2$ -step and  $k_2^2$ -step become too unstable to exist as intermediates, then the formation of  $T_2^0$  and  $T_N^+$  involves transition states  $TS_3$  and  $TS_4$ , respectively. The  $pK_a$  of conjugate acid of leaving group is significantly larger in  $k_4^2$ -step than that in  $k_{-1}^2$ -step and consequently  $k_{-1}^2 \gg k_4^2$  which leads to  $k_4^2$ -step as the rate-determining step. Similarly, the  $pK_a$  of conjugate acid of leaving group is larger in  $k_{-2}^2$ -step than that in  $k_3^2$ -step and this leads to  $k_3^2 \gg k_{-2}^2$  which in turn reveals  $k_3^2$ -step as the rate-determining step. The observed rate law:  $\text{rate} = k_{\text{obs}} [\mathbf{3}]_T$  and Scheme 2 give Eq. (3)

$$k_{\text{obs}} = \frac{k_1^2 [\text{H}_2\text{O}] + k_2^2 [\text{H}_2\text{O}] K_{\text{eq}}^2 [\text{H}^+]}{1 + K_{\text{eq}}^2 [\text{H}^+]} \quad (3)$$



Scheme 2. Structures  $T_2^\pm$ ,  $T_0^+$ ,  $TS_3$ ,  $TS_4$ .

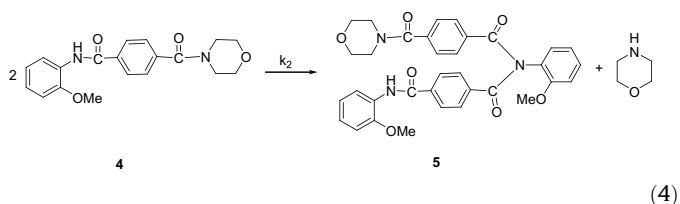
where  $k_1^2$  and  $k_2^2$  represent second-order rate constants for uncatalyzed and specific acid-catalyzed hydrolysis of **3**. Empirical Eq. (1) is similar to Eq. (3) with  $k_0 = k_1^2 [\text{H}_2\text{O}]$ ,  $k_c = k_2^2 [\text{H}_2\text{O}]$  and  $K = K_{\text{eq}}^2$ .

It is evident from Schemes 1 and 2 that  $K_{\text{eq}}^1 = 1/K_a^{1\text{H}^+}$  and  $K_{\text{eq}}^2 = 1/K_a^{3\text{H}^+}$  where  $K_a$  represents concentration ionization constant. The calculated values of  $K$  ( $= K_{\text{eq}}^1$  or  $K_{\text{eq}}^2$ ) were used to calculate  $K_a$  for  $1\text{H}^+$  and  $3\text{H}^+$  which are summarized in Table 1. The value of thermodynamic  $pK_a$  ( $= -1.17$ ) for protonated *N,N*-dimethylbenzamide, determined spectrophotometrically in the absence of an inert salt [9], may be compared with concentration  $pK_a$  ( $= -0.60$ ) for  $1\text{H}^+$ . The value of concentration  $K_a$  for  $1\text{H}^+$  is  $\sim 3$ -fold larger than that for  $3\text{H}^+$  at  $1.0$  M ionic strength (Table 1) which could be partly attributed to steric hindrance due to 2- $\text{CONHC}_6\text{H}_4\text{OMe}$ -2' group in  $1\text{H}^+$  [14]. The value of  $K_a$  for  $1\text{H}^+$  at  $1.0$  M ionic strength is  $\sim 1.3$ - to  $1.8$ -fold larger than that at  $5.0$  M ionic strength (Table 1). Similarly, the calculated concentration  $K_a$  ( $= 1.23$  M) value for benzamide at  $6.0$  M ionic strength (by LiCl), calculated from kinetic data of Ref. [20], is  $\sim 28$ - to  $117$ -fold smaller than thermodynamic  $K_a$  ( $= 34$ ,  $55$ ,  $62$  and  $144$  M) obtained spectrophotometrically in the absence of an inert salt [9]. Various reports show that the values of  $pK_a$  of O-protonated amide groups vary in the range of  $0$  to  $-3$  [21].

In order to assess the rate enhancement due to intramolecular-ity of the reaction as displayed by Scheme 1, one requires the value

<sup>1</sup> We thank the reviewer for suggesting this point.

of second-order rate constant ( $k_2$ ) for the bimolecular reaction (Eq. (4)) carried out under the experimental condition of present study.



But the formation of product **5** from **4** is apparently impossible for the reason that the rate of hydrolysis of **4** should be much faster than that of imide (**5**) formation from **4** under the present experimental conditions. This speculation is based upon, at least, following two reasons. (i) Although the basicity of amide nitrogen of **4** may not be significantly different from that of  $\text{H}_2\text{O}$  in terms of  $\text{p}K_b$  values, the nucleophilic site of **4** is more sterically hindered than that of  $\text{H}_2\text{O}$ . (ii) The concentration of  $\text{H}_2\text{O}$  is  $\sim 2.7 \times 10^5$ -fold larger than that of **4**. Thus, an underestimated value of rate enhancement of  $2 \times 10^6$ -fold due to intramolecular carboxamide group-assisted cleavage of the amide bond in **1** may be obtained by comparing the value of  $k_c$  for  $1\text{H}^+$  and  $k_c/[\text{H}_2\text{O}]$  for  $3\text{H}^+$  (Table 1). It is perhaps noteworthy that the value of  $k_0$  ( $=2.19 \times 10^{-5} \text{ s}^{-1}$ ) and the effective molarity ( $=2 \times 10^6 \text{ M}$ ) due to intramolecularity in the acidic aqueous cleavage of **1** may give  $k_0/[\text{H}_2\text{O}]$  as  $1.1 \times 10^{-11} \text{ M}^{-1} \text{ s}^{-1}$  ( $=2.19 \times 10^{-5} \text{ s}^{-1}/2 \times 10^6 \text{ M}$ ) for **3** provided  $k_c/k_0$  value remained same for the aqueous cleavage of both **1** and **3**. Thus, the estimated value of  $k_0 = 6 \times 10^{-10} \text{ s}^{-1}$  for uncatalyzed hydrolysis of **3** is similar to the corresponding reported  $k_0$  value for uncatalyzed hydrolysis of formamide [11].

#### 4. Conclusions

Perhaps this is the first report which reveals the rate enhancement of much more than  $10^6$ -fold due to intramolecular carboxamide group assistance in the cleavage of the amide bond in mild acidic aqueous solution. Intermolecular specific acid catalytic component is  $\sim 500$ -fold. The present finding predicts an efficient conversion of phthalamide, *N*-substituted, *N,N'*-disubstituted and *N,N,N'*-trisubstituted phthalamides to the corresponding phthalamide and *N*-substituted phthalamides under mild acidic pH. The finding of large rate enhancement in the intramolecular amide nitrogen-assisted cleavage of an amide bond under neutral/mild acidic pH reveals the possibility that the acidic hydrolysis of a peptide can be assisted by a neighboring amide group, which could be relevant to the action of numerous enzymes.

#### Acknowledgments

The authors thank the Ministry of Science, Technology and Innovation for ScienceFund (Project No.: 14-02-03-4014) and the University of Malaya for financial support.

#### Appendix A. Supplementary data

Fig. 1, Experimental details for synthesis of **1**, **3**, kinetic measurements, product identification and NMR spectra for **1**. Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.bioorg.2008.03.003.

#### References

- [1] J.A. Shafer, H. Morawetz, *J. Org. Chem.* 28 (1963) 1899–1901.
- [2] (a) W.P. Jencks, *Catalysis in Chemistry and Enzymology*, McGraw-Hill, New York, 1969; (b) T.C. Bruice, S.J. Benkovic, *Bioorganic Mechanisms*, W.A. Benjamin, New York, 1966; (c) M.L. Bender, *Mechanisms of Homogeneous Catalysis from Protons to Proteins*, Wiley, New York, 1971; (d) T.H. Fife, *Adv. Phys. Org. Chem.* 11 (1975) 1–122; (e) A.J. Kirby, *Adv. Phys. Org. Chem.* 17 (1980) 183–278; (f) W.P. Jencks, *Adv. Enzymol.* 43 (1975) 219–410; (g) A.R. Ferscht, *Enzyme Structure and Mechanism*, Freeman, New York, 1977.
- [3] T.H. Fife, in: E.E. Van Tamelen (Ed.), *Bioorganic Mechanisms*, Academic Press, New York, 1977, p. 93, Chapter 5.
- [4] (a) A.F. Hegarty, T.C. Bruice, *J. Am. Chem. Soc.* 92 (1970) 6575–6588; (b) A.J. Kirby, T.G. Mujahid, P. Camilleri, *J. Chem. Soc. Perkin Trans. 2* (1979) 1610–1616; (c) T.H. Fife, N.W. Duddy, *J. Am. Chem. Soc.* 105 (1983) 74–79; (d) M.N. Khan, *J. Chem. Soc. Perkin Trans. 2* (1988) 213–219; (e) T.H. Fife, L. Chaffee, *J. Org. Chem.* 65 (2000) 3579–3586; (f) T.H. Fife, R. Singh, R. Bemb, *J. Org. Chem.* 67 (2002) 3179–3183.
- [5] T.H. Fife, B.R. DeMark, *J. Am. Chem. Soc.* 99 (1977) 3075–3080.
- [6] T. Cohen, J. Lipowitz, *J. Am. Chem. Soc.* 86 (1964) 5611–5616.
- [7] (a) M.L. Bender, *J. Am. Chem. Soc.* 79 (1957) 1258–1259; (b) M.L. Bender, Y.-L. Chou, F. Chloupek, *J. Am. Chem. Soc.* 80 (1958) 5380–5384; (c) A.J. Kirby, P.W. Lancaster, *J. Chem. Soc. Perkin Trans. 2* (1972) 1206–1214; (d) M.N. Khan, A. Ariffin, *Org. Biomol. Chem.* 1 (2003) 1404–1408; (e) A.M. Granados, R.H. deRossi, *J. Org. Chem.* 66 (2001) 1548–1552; (f) F.M. Menger, M. Ladika, *J. Am. Chem. Soc.* 110 (1988) 6794–6796; (g) R. Kluger, J.C. Hunt, *J. Am. Chem. Soc.* 111 (1989) 5921–5925.
- [8] Y.-L. Sim, A. Ariffin, M.N. Khan, *J. Org. Chem.* 72 (2007) 2392–2401.
- [9] C.A. Bunton, S.J. Farber, A.J.G. Milbank, C.J. O'Connor, T.A. Turney, *J. Chem. Soc. Perkin Trans. 2* (1972) 1869–1875.
- [10] J.W. Barnett, C.J. O'Connor, *J. Chem. Soc. Perkin Trans. 2* (1973) 220–222.
- [11] J. Hine, R.S.-M. King, W.R. Midden, A. Sinha, *J. Org. Chem.* 46 (1981) 3186–3189.
- [12] W.P. Jencks, *Chem. Soc. Rev.* 10 (1981) 345–375.
- [13] P.Y. Bruice, *Organic Chemistry*, Prentice-Hall, Inc., USA, New Jersey, 1995, pp. A-7, A-8, Appendix II.
- [14] J. Hine, *Structural Effects on Equilibria in Organic Chemistry*, John Wiley & Sons, New York, 1975, pp. 66.
- [15] C.J. Perry, Z. Parveen, *J. Chem. Soc. Perkin Trans. 2* (2001) 512–521.
- [16] M.D. Hawkins, *J. Chem. Soc. Perkin Trans. 2* (1976) 642–647.
- [17] (a) C.J. O'Connor, *Quart. Rev. Chem. Soc.* 24 (1970) 553–564; (b) A. Williams, *J. Am. Chem. Soc.* 98 (1976) 5645–5651. and references therein.
- [18] D. Zahn, *J. Phys. Chem. B* 107 (2003) 12303–12306.
- [19] M. Novak, G.A. Bonham, L.K. Mohler, K.M. Peet, *J. Org. Chem.* 53 (1988) 3903–3908.
- [20] (a) M.N. Khan, *J. Chem. Soc. Perkin Trans. 2* (1988) 1129–1134; (b) M.N. Khan, *J. Chem. Soc. Perkin Trans. 2* (1990) 435–444.
- [21] R.S. Brown, in: A. Greenberg, C.M. Breneman, J.F. Liebman (Eds.), *The Amide Linkage: Structural Significance in Chemistry, Biochemistry, and Materials Science*, John Wiley & Sons, Inc., Hoboken, New Jersey, 2003, Chapter 4, and references therein.